

Effect of Inhaled Interleukin-5 on Activity and Number of Eosinophils in Circulation from Asthmatics*

吸入白细胞介素-5对支气管哮喘患者外周血嗜酸性粒细胞数及其活化状态的影响

Xie Zhengfu Liu Guangnan Qin Shouming Den Jiazen Shi Huanzhong Xu Hui
谢正福 柳广南 覃寿明 邓家珍 施焕中 许辉

(Department of Internal Medicine, First Affiliated Hospital, Guangxi Medical University, 6 Binhulu, Nanning, Guangxi, 530021)
(广西医科大学第一附属医院内科 南宁市滨湖路6号 530021)

Abstract For examining the effects of recombinant human(rh) IL-5 inhalation on changes of activity and number of circulating eosinophils in allergic asthmatics, as well as concentrations of serum IgE, a randomized double-blind, placebo-controlled study design was employed, in which each subject acted as his or her own control. Eight nonsmoking patients with allergic asthma were administered rhIL-5 by nebulization, total blood nuclear cell counts and differentials, as well as concentrations of ECP and IgE in serum were determined before, and at 2 h, 24 h, 48 h after inhalation, respectively. Eosinophil numbers and ECP levels within the control group did not appear to change from baseline at any time throughout the study, eosinophil numbers from baseline($3.6 \pm 1.8 \times 10^5 / \text{mL}$) to $6.3 \pm 1.2 \times 10^5 / \text{mL}$ ($P < 0.01$) at 24 h, and to $5.7 \pm 0.9 \times 10^5 / \text{mL}$ ($P < 0.01$) at 48h after IL-5 inhalation. Accompanying this significant blood eosinophilia was a significant elevation of serum ECP levels. Compared with baseline value ($6.3 \pm 1.1 \text{ pg/mL}$), inhalation of IL-5 lead to ECP levels increase with time, reaching $17.6 \pm 2.8 \text{ pg/mL}$ at 24 h ($P < 0.01$); and this elevated ECP levels lasted at least 48 h ($18.1 \pm 2.9 \text{ pg/mL}$, $P < 0.01$). IL-5 inhalation had no significant effect of levels of serum total IgE. our findings provided direct evidence that IL-5 not only induced a significant blood eosinophilia, but also resulted in the activation of circulating eosinophils.

Key words Asthma, eosinophil cationic protein, Eosinophil, IL-5

摘要 随机选择 8 例哮喘患者雾化吸入人重组 IL-5, 分别吸入前、吸入后 2 h、24 h 和 48 h 进行外周血有核细胞计数和分类, 以放射免疫法测定血清中嗜酸性阳离子蛋白 (ECP) 及总 IgE 水平。结果, 吸入 IL-5 后嗜酸性粒细胞 (EOS) 占细胞总数的百分比, EOS 绝对细胞数以及 ECP 水平均随时间而明显升高, 至 24 h 达最高值, 48 h 后仍维持在较高水平; IL-5 在整个实验过程中对血清总 IgE 水平无明显影响。表明, 吸入 IL-5 不仅可以促使循环中 EOS 数明显增多, 且还能招致其活化从而参与哮喘的发病过程。

关键词 支气管哮喘 嗜酸性阳离子蛋白 嗜酸性粒细胞 白细胞介素-5

中图分类号 R 256.25

Bronchial asthma, even mild asthma, is associated with persistent inflammation of the bronchial mucosa characterized by infiltration of eosinophils^[1-4]. This inflammation has been suggested to be responsible for airway hyperreactivity^[5]. One of these eosinophilic proteins, eosinophil cationic protein (ECP), has been documented to be increased in the airway and circulation in asthma^[6-7]. Furthermore,

both circulating eosinophil numbers and serum ECP levels and their interrelationship may be of value in assessing the severity of asthma^[8].

A number of cytokines with selective actions on eosinophils have been identified, including interleukin (IL)-3, IL-5, and granulocyte-macrophage colony-stimulating factor. One of the most important is IL-5 which promotes terminal differentiation of the committed eosinophil precursor^[9] as well as enhancing the effector capacity of mature eosinophils^[10]. IL-5 also prolongs the survival of eosinophil in vitro^[11], and selectively enhances eosinophil degranulation, antibody-dependent cytotoxicity^[12]. It has been demonstrated that IL-5 gene expression in circulating CD4⁺ cell and

1998-05-08收稿

* Supported in part by a grant No. 96053 from the Education Commission of Guangxi Zhuang Autonomous Region, P. R. C., and in part by a research grant No. 9532012 from Science and Technology Commission of Guangxi Zhuang Autonomous Region.

serum ECP concentrations were both significantly increased in asthma, particularly in acute severe asthma, moreover, significant correlations were observed between IL-5 expression and ECP levels^[13]. By topical instillation of recombinant human (rh) IL-5 into the lower airways, we recently indicated that IL-5 is capable of increasing ECP levels in bronchoalveolar lavage (BAL) fluid in allergic asthmatics^[14]. The present study was conducted to investigate the effects of inhaled rhIL-5 on changes of numbers of eosinophils in circulation, as well as concentrations of ECP and total IgE in serum in patients with allergic bronchial asthma.

2 Methods

2.1 Subjects

Eight nonsmoking asthmatics were enrolled in this study (Table 1). Asthma was defined as a clinical history of intermittent chest tightness, wheeze, cough, or shortness of breath, and documented reversible airflow limitation either spontaneously or with treatment in the preceding year (20% change in FEV₁ or peak expiratory flow PEF). All patients had mild atopic asthma, with baseline forced expiratory volume in 1 s (FEV₁) greater than 70% of predicted value, requiring only intermittent use of inhaled β_2 -agonists. All patients had a provocative concentration of methacholine producing a 20% fall in FEV₁ (PC₂₀-Mch) < 8 mg/mL. Each patient had one or more documented positive skin prick test responses to aeroallergens. None had received inhaled or oral corticosteroids in the previous 3 months. Any subjects with a history suggestive of a viral upper respiratory tract infection within the 4 weeks preceding or during the study were excluded.

The study protocol was approved by the Ethics Committee of Guangxi Medical University, P. R. China, and all subjects provided written consent.

Table 1 Characteristics of the asthmatic subjects

Subject No.	Age (a)	Sex	Atopy	IgE (IU/mL)	FEV ₁ % predicted	PC ₂₀ -Mch (mg/mL)
1	58	M	+	199	97	1.27
2	38	M	+	520	108	0.09
3	43	M	+	178	89	0.17
4	39	F	+	442	96	2.53
5	27	F	+	248	110	5.12
6	19	M	+	968	82	0.62
7	42	M	+	113	103	2.58
8	35	F	+	104	99	1.26
Mean \pm SEM		38 \pm 4		347 \pm 103	98 \pm 3	0.92 \pm 1.64

* Geometric mean \pm geometric SEM.

2.2 Study design

A randomized double-blind, placebo-controlled study design was employed, in which each subject acted as his or her own control.

At a preliminary visit, a full history and examination was performed and skin prick tests to a panel of

common aeroallergens were carried out. At the same time, blood was taken for measuring total nuclear blood cells and cell differentials, and concentrations of ECP and total IgE; methacholine challenge test was also done. The following afternoon (24h after baseline measurements) 10 μ g of rhIL-5 (Genzyme Co., Boston, MA) in vehicle (0.1% bovine serum albumin in 0.9% saline) or vehicle only was inhaled as a 0.5 mL nebulized solution, the chamber was refilled twice with 0.5 mL vehicle, and the nebulization was kept running to scavenge any remaining rhIL-5. The dose of rhIL-5 was based upon a preliminary study involving two asthmatic patients.

Blood cytology, concentrations of ECP and total IgE were measured at 2 h, 24 h, and 48 h after the inhalation of rhIL-5 or vehicle. At least 4 weeks were allowed to elapse between the two inhalation, and the order of inhalation of rhIL-5 or vehicle was randomized.

2.3 Blood samples

Venous blood samples were obtained from each subject. Samples were collected in either ethylenediaminetetraacetic acid (EDTA)-treated tubes for total and differential blood cell counts or untreated tubes to obtain sera for determining ECP and total IgE levels, and sera were stored at -70°C for later determination. Total cells were counted with a hemocytometer and different cell counts were made from blood smears stained with Diff-Quik Staining (Sigma Chemical Company, St. Louis, Mo). Blood smears were counted blind in coded random order by two independent observers, using a light microscope. A total of 300 cells were enumerated for differential cell counts in identifying monocytes, neutrophils, lymphocytes, eosinophils and basophils, and absolute numbers of each cell type were calculated. The interobserver varied < 8%.

2.4 ECP assay

Serum samples stored at -70°C were thawed. Levels of ECP in serum were determined with ECP Fluoroimmunoassay kits (Pharmacia AB, Uppsala, Sweden) on the procedures recommended by the manufactures, with result expressing in pg/mL. Assay sensitivity for ECP was 2 pg/mL.

2.5 Total IgE assay

Concentrations of total IgE in serum were determined with total IgE enzyme immunoassay test kits (Golden Bridge International Inc., WA) on the procedures recommended by the manufactures, with result expressing in IU/mL. The minimum detectable concentration of IgE by this assay was established to be 5 IU/mL.

2.6 Statistical analysis

All data were presented as mean \pm standard error of mean (SEM). Measures variance (ANOVA) for data were conformed to be in a normal distribution, to be a nonparametric distribution by Friedman's test (confirmed by the Shapiro-Wilk W test). Paired *t* test was used to compare data obtained at the same time

points between the two inhalations.

3 Results

3.1 Effect of inhaled IL-5 on blood cytology

All subjects tolerated the procedures without complication. The effects of rhIL-5 inhalation on changes of blood cell population and cell differential counts were shown in Table 2 and Figure 1. In this study, vehicle challenge was not able to cause any changes in total cell numbers, percentages of each cell type, or absolute numbers of each cell type including eosinophils in peripheral blood (all $P < 0.05$). Blood total cell numbers, as well as percentages of monocytes, neutrophils and lymphocytes were also non-significantly increased at three occasions after IL-5 inhalation compared with baseline (all $P > 0.05$). However, there was an increase in percentages of circulating eosinophils after IL-5 inhalation, which increased with time as significantly comparing to baseline value or vehicle controls, respectively, reaching a maximum at 24 h, and lasted less than 48 h (all $P < 0.01$). Moreover, a similar increase in absolute numbers of eosinophils in blood after IL-5 inhalation was observed.

Table 2 Comparison of total cell counts and differentials in peripheral blood from allergic asthmatics challenged with interleukin-5 and vehicle control

Group	Total cells ($\times 10^5$ /mL)	Cell differentials (%)			
		Monocytes	Neutrophils	Lymphocytes	Eosinophils
Interleukin-5					
Baseline	66.3 ± 5.7	1.9 ± 0.6	65.7 ± 2.1	27.0 ± 2.3	5.4 ± 1.5
2h	69.3 ± 5.4	1.5 ± 0.5	65.3 ± 2.2	28.1 ± 1.9	5.1 ± 1.2
24h	68.9 ± 5.8	1.8 ± 0.7	64.5 ± 3.6	24.8 ± 2.4	9.0 ± 1.5†
48h	68.1 ± 4.6	2.0 ± 0.7	61.3 ± 3.6	28.0 ± 2.5	8.6 ± 1.5†
Vehicle					
Baseline	66.3 ± 4.9	2.0 ± 0.2	64.3 ± 2.0	28.8 ± 1.7	5.0 ± 1.2
2h	66.9 ± 5.3	1.8 ± 0.8	64.6 ± 1.9	28.4 ± 1.4	5.2 ± 1.3
24h	67.3 ± 4.1	0.8 ± 0.5	65.4 ± 2.2	27.8 ± 1.7	5.1 ± 1.3
48h	66.5 ± 4.5	1.6 ± 0.5	63.3 ± 2.8	29.9 ± 2.7	5.4 ± 1.2

* Data were presented as mean ± SEM ($n = 8$ for each group).

† Compared with baseline or vehicle controls. $P < 0.01$.

3.2 Effects of inhaled IL-5 on ECP levels in serum

Baseline measurements of ECP before both challenge showed no significant difference in serum from allergic asthmatics (6.3 ± 1.1 pg/mL with IL-5 and 6.8 ± 1.2 pg/mL with vehicle, respectively, $P > 0.05$) (Figure 2). Concentrations of serum ECP within the vehicle group had no change baseline throughout the study (all $P > 0.05$). It meant vehicle inhalation of asthmatics did not lead to elevations of ECP levels in circulation. Serum ECP levels (6.8 ± 1.3 pg/mL) at 2 h after IL-5 inhalation were still not different in baseline value ($P > 0.05$). ECP levels increased from baseline to a higher extent at 24 h (17.8 ± 2.8 pg/mL, $P < 0.01$) and at 48 h (18.1 ± 2.9 pg/mL, $P < 0.01$) after IL-5 inhalation.

No significant difference in concentrations of ECP between IL-5 and vehicle inhalation was observed at 2 h. However, ECP concentrations at 24 h and 48 h af-

ter IL-5 inhalation were significantly higher as compared with their vehicle controls, respectively.

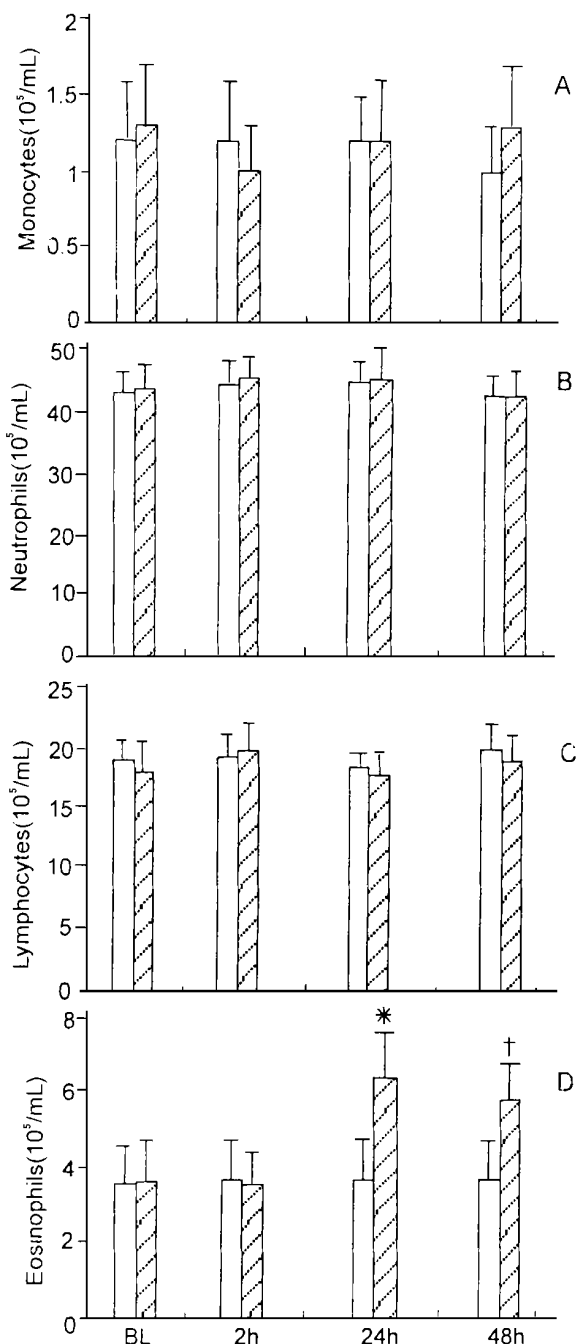


Fig. 1 Comparison of numbers of each type cell in blood from eight allergic asthmatics challenged with recombinant human IL-5 (hatched) or vehicle (white). BL= baseline. * Compared with baseline measurement or vehicle controls, $P < 0.01$. Panel A: monocytes; Panel B: neutrophils; Panel C: lymphocytes; Panel D: eosinophils.

3.3 Effects of inhaled IL-5 on total IgE levels in serum

As shown in Figure 3, baseline total IgE in serum before both challenges showed no significant difference. Neither rhIL-5 nor vehicle inhalation made serum IgE levels change throughout the study (all $P >$

4 Discussion

Previous studies have demonstrated inverse relationships between blood eosinophil numbers and FEV₁

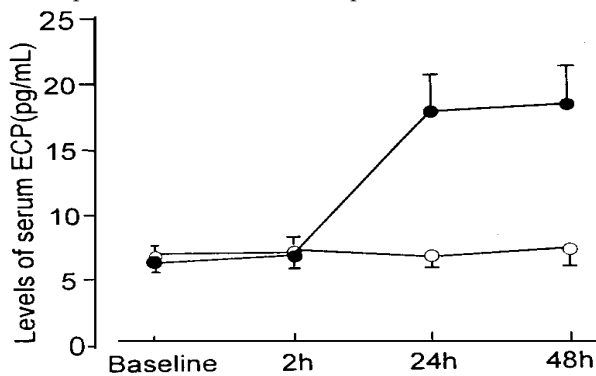


Fig. 2 Comparison of ECP levels in serum from eight allergic asthmatics challenged with recombinant human IL-5 or vehicle. * Compared with baseline measurement or vehicle controls, $P < 0.01$ —○— Vehicle; —●— IL-5.

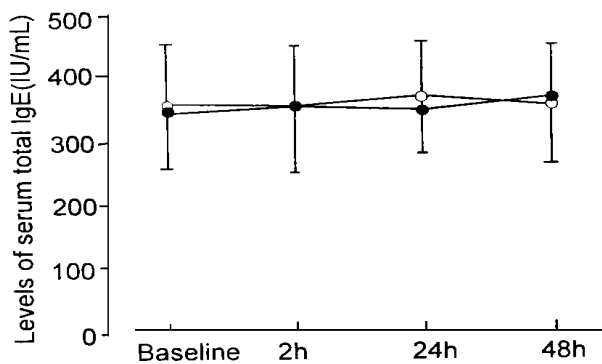


Fig. 3 Comparison of total IgE levels in serum from eight allergic asthmatics challenged with recombinant human IL-5 or vehicle. —○— Vehicle; —●— IL-5.

and airway responsiveness^[15,16] in asthmatics. Serum ECP levels have been demonstrated to be more closely related to the lung function during the late asthmatic reaction than the number of circulating eosinophils, suggesting that the activity of the eosinophils might be more important for the development of the late asthmatic reaction than the number^[17-19]. Griffin and coworkers' study indicated that in patients' with asthma and pronounced eosinophilia, the lung function of the patients was principally related to the number of circulating eosinophils, whereas, when their eosinophilia was reduced to moderate levels, the patients' lung function was closer related to the activity of the eosinophils^[20]. Serum from asthmatics, in comparison with the serum of healthy subjects, has a higher level of IL-5^[21,22]. And serum concentrations of IL-5 have been reported to be related with circulating eosinophil numbers^[23,24]. Lai and colleagues have demonstrated that IL-5 gene expression in circulating CD4 cells and serum ECP concentrations were both significantly increased in asthma, particularly in acute severe asthma, moreover, significant correlations were

observed between IL-5 expression and ECP levels^[13]. Actually, IL-5 is of particular interest in the pathophysiology of asthma as it is associated with eosinophilic inflammation.

The ability of IL-5 to induce an increase in number of circulating eosinophils has been previously demonstrated in various animal models by several groups^[25,26].

We have found that the total eosinophils (BMK-13 cells) and the activated eosinophils (EG2 cells) in bronchial mucosa, as well as the eosinophil numbers in BAL fluid from IL-5-challenged segments were significantly higher than those in vehicle-challenged segments^[14]. Eosinophil activation assessed by secretion of ECP was also increased significantly in bronchial mucosa and BAL fluid. The results strongly suggested that IL-5 was capable of inducing eosinophil infiltration into the asthmatic airways, as well as the activation of infiltrating eosinophils. In the present study, our data extend these findings by observing the effects of rhIL-5 challenge on the number of circulating eosinophils and concentrations of serum ECP in allergic asthmatics.

In this study, we observed that allergic asthmatics had a proportion of blood eosinophils and serum ECP concentrations even before rhIL-5 challenge. rhIL-5 inhalation had no effect on the changes of blood total cells and percentages of monocytes, neutrophils, and lymphocytes. In addition, a few basophils could be seen in all blood smears studied, no increase in circulating basophils was observed even after rhIL-5 inhalation. Of particular interest was the eosinophils increases in circulation of asthmatic patients challenged with rhIL-5. The changes were time-course-related, with a most response at 24 h and last less than 48 h. Both the percentage and the absolute number of circulating eosinophils were significantly higher at 24 h and 48 h after rhIL-5 challenge than baseline measurement before challenge. We also found that serum ECP levels were elevated in asthmatics by rhIL-5 inhalation in a similar manner. Our results demonstrated that inhalation of 10⁴ g rhIL-5 in patients with allergic asthma was able to not only increase eosinophils in circulation, but also result in activation of increased circulating eosinophils. In addition, in the same study design, we have also found that rhIL-5 challenge could increase airway responsiveness in patients with allergic bronchial asthma, accompanying a significant eosinophilia and elevated concentrations of ECP in induced sputum (these findings would be reported separately).

The exact mechanism by which inhaled IL-5 contribute to blood eosinophilia is still unknown. It seems likely that after it is administered topically into the airways, IL-5 penetrates into circulation through bronchial mucosa as well as blood microvessel wall, and then exerts its possible systemic hormone-like ac-

tions. Collins and coworkers have reported^[27] that intravenous injection of IL-5 of guinea pigs stimulated a significant increase in the numbers of circulating eosinophils, and this increase in circulating eosinophils corresponded with a reduction in the number of displaceable eosinophils recovered after flushing out the femur bone marrow cavity, suggesting that IL-5 acts as a hormone to stimulate the release into the circulation of a rapidly mobilizable pool of bone marrow eosinophils. Now that our present study provided important evidence that IL-5 produced in the airways may have systemic hormone-like actions, we speculated that IL-5 was capable of stimulating the release of a pool of mature eosinophils from the bone marrow into the circulation in a similar manner. On the other hand, IL-5 might be able to increase eosinophil differentiation from bone marrow precursor cells, since IL-5 possesses this ability^[9].

Immunoglobulin isotype switching to IgE synthesis is mainly mediated by IL-4 and costimulatory signals provided by CD4⁺ T cells^[28]. IL-5 has been reported to enhance IL-4-dependent IgE lproduction^[29]. When C57BL/6 mice were challenged with antigen twice a week for 2 weeks, Chu et al.^[30] found that lung tissue IL-5 and IL-4 expressing cells increased in number, which was similar to the profile of serum total IgE production. And both IL-5 and IL-4 expressing cells correlated with serum total IgE levels. This suggests that IL-5, along with IL-4, could be a major modulator of IgE production in this sensitized mouse model. In another animal experiment^[31], Coffman and colleagues have demonstrated that when BALB/c were infected with the nematode *Nippostrongylus brasiliensis*, large numbers of eosinophils appeared in their blood and lungs and their serum IgE was increased; injection of an anti-IL-5 monoclonal antibody completely suppressed the blood eosinophilia and the infiltration of eosinophils in the lungs of parasitized mice but had no effect on serum IgE; in contrast, an antibody to IL-4 inhibited parasite-induced IgE but not the eosinophilia. These results indicate that IL-5 is important in eosinophil production in vivo and that IgE production is regulated by IL-4, not by IL-5. This inability of anti-IL-5 to inhibit the IgE response is consistent with the findings in our present study, which showed that rhIL-5 inhalation of atopic asthmatics was not able to cause any changes in serum IgE levels in the study, although a significant blood eosinophilia and a marked elevation of serum ECP concentrations were observed.

Eosinophils are known to secrete a number of basic proteins, including major basic protein (MBP), ECP, Eosinophil-derived neuroxin, and eosinophil peroxidase that have profound effects on airway cells^[1,32]. Release of these products is associated with increases in vascular permeability, bronchoconstriction, and destruction of airway epithelial cells^[1].

In particular, MBP has been shown to cause airway constriction and airway hyperresponsiveness^[33,34]. In addition, eosinophil activation also results in the release of a number of important lipid mediators, including leukotriene C₄, which can contract airway smooth muscle, and platelet-activating factor, which can contract airway smooth muscle as well as increase bronchial responsiveness^[35]. Thus, eosinophils possess properties that can directly or indirectly cause airway obstruction and promote bronchial hyperreactivity. Since IL-5 is capable of increasing circulating eosinophils, as well as the activation of eosinophils, it is possible that IL-5 exerts its biological effects in the pathogenesis of asthma by means of a role for eosinophils. Therefore, drugs that interfere with IL-5 synthesis or IL-5 receptor antagonists could be beneficial in the treatment of asthma.

Acknowledgments

The authors are grateful to Professor Jun-fu Xiong, Zhi-xing Liao, and Bin-bin Liao for their valuable participation in this study.

References

- Gleich G J. The eosinophil and bronchial asthma: current understanding. *J Allergy Clin Immunol*, 1990, 85: 422~436.
- Holgate S T, Roche W R, Church M K. The role of eosinophil in asthma. *Am Rev Respir Dis*, 1991, 143: S66~S70.
- Oddera S, Silverstri M, Balbo A et al. Airway eosinophilic inflammation, epithelial damage, and bronchial hyperresponsiveness in patients with mild-moderate, stable asthma. *Allergy*, 1996, 51: 100~107.
- Synek M, Beasley R, Frew A J et al. Cellular infiltration of the airways in asthma of vary severity. *Am J Respir Crit Care Med*, 1996, 154: 224~230.
- Hargreave F E. Late phase asthmatic responses and airway inflammation. *J Allergy Clin Immunol*. 1989, 83: 525~527.
- Robinson D S, Assoufi B, Durham S R et al. Eosinophil cationic protein (ECP) and eosinophil protein (EPX) concentrations in serum and bronchial lavage fluid in asthma. Effect of prednisolone treatment. *Clin Exp Allergy*, 1995, 25: 1118~1127.
- Oosterhoff Y, Kauffman H F, Rutgers B et al. Inflammation cell number and mediators in bronchial lavage fluid and peripheral blood in subjects with asthma with increased nocturnal airways narrowing. *J Allergy Clin Immunol*, 1995, 96: 219~229.
- Bjornsson E, Janson C, Hakansson L et al. Serum eosinophil cationic protein in relation to bronchial asthma in a young Swedish population. *Allergy*, 1994, 49: 730~736.
- Clutterbuck E J, Hrist M A, Sanderson C J. Human interleukin-5 (IL-5) regulates the production of eosinophils in human bone marrow cultures: comparison and interaction with IL-1, IL-3, IL-6, and GM-CSF. *Blood*, 1988, 73: 1504~1513.
- Lopez A F, Sanderson C J et al. 1988. Recombinant human interleukin-5 is a selective activator of human eosinophil function. *J Exp Med*, 1988, 167: 219~224.
- Yamaguchi Y, Hayashi Y, Sugama Y et al. Highly purified murine interleukin-5 (IL-5) stimulates eosinophil

- function and prolongs in vitro survival. *J Exp Med*, 1988, 167: 1734~ 1742.
- 12 Fujisawa T, Abu-Ghazaleh R, Kita H et al. Regulatory effect of cytokines on eosinophil degranulation. *J Immunol*, 1990, 144: 642~ 646.
 - 13 Lai C K W, Ho A S S, Chan C H S et al. Interleukin-5 messenger RNA expression in peripheral blood CD4⁺ cells in asthma. *J Allergy Clin Immunol*, 1996, 97: 1320~ 1328.
 - 14 Shi H Z, Qin S M, Huang G W et al. Infiltration of eosinophils into the asthmatic airways caused by interleukin-5. *Am J Respir Cell Mol Biol*, 1997, 16: 220~ 224.
 - 15 Horn B R, Robin E D, Theodore J, Van Kessel A. Total eosinophil counts in the management of bronchial asthma. *N Engl J Med*, 1975, 292: 1152~ 1155.
 - 16 Taylor K J, Luksza A R. Peripheral blood eosinophil counts and bronchial responsiveness. *Thorax*, 1987, 42: 452~ 456.
 - 17 Venge P, Dahl R, Peterson C G B. Eosinophil granule proteins in serum after allergen challenge of asthmatic patients and the effects of anti-asthmatic medication. *Int Arch Allergy Appl Immunol*, 1988, 87: 306~ 312.
 - 18 Venge P, Dahl R. Are blood eosinophil number and activity important for the development of the late asthmatic reaction after allergen challenge? *Eur Respir J*, 1989, 2 (Suppl 6): 430s~ 434s.
 - 19 Rak S, Liwhagen O, Venge P. The effect of immunotherapy on bronchial hyperresponsiveness and eosinophil cationic protein in pollen-allergic patients. *J Allergy Clin Immunol*, 1988, 82: 470~ 480.
 - 20 Griffin E, Håkansson L, Formgren H et al. Blood eosinophil number and activity in relation to lung function in patients with asthma and with eosinophilia. *J Allergy Clin Immunol*, 1991, 87: 548~ 557.
 - 21 Alexander A G, Barkans J, Moqbel R et al. Serum interleukin-5 concentrations in atopic and non-atopic patients with glucocorticoid-dependent chronic severe asthma. *Thorax*, 1994, 49: 1231~ 1233.
 - 22 Corrigan C J, Haczku A, Gemou-Engesaeth V et al. CD4⁺ T-lymphocyte activation in asthma is accompanied by increased serum concentrations of interleukin-5. Effect of glucocorticoid therapy. *Am Rev Respir Dis*, 1993, 147: 540~ 547.
 - 23 Koike T, Enokihara H, Arimura H et al. Serum concentrations of IL-5, GM-CSF, and IL-3 and the production by lymphocytes in various eosinophilia. *Am J Hematol*, 1995, 50: 98~ 102.
 - 24 Koike T, Enokihara H, Arimura H et al. Serum concentrations of IL-5 and the production by lymphocytes in reactive eosinophilia. *Int Arch Allergy Immunol*, 1995, 108 (suppl 1): 16~ 18.
 - 25 Foster P S, Hogan S P, Ramsay A J et al. Interleukin 5 deficiency abolishes eosinophilia, airways hyperreactivity, and lung damage in a mouse asthma model. *J Exp Med*, 1996, 183: 195~ 201.
 - 26 Portanova J P, Christine L J, Rangwala S H et al. Rapid and selective induction of blood eosinophilia in guinea pigs by recombinant human interleukin-5. *Cytokine*, 1995, 7: 775~ 783.
 - 27 Collins P D, Marleau S, Griffiths-Johnson D A et al. Cooperation between interleukin-5 and chemokine eotaxin to induce eosinophil accumulation in vivo. *J Exp Med*, 1995, 182: 1169~ 1174.
 - 28 Kopf M, Le Gros G, Bachmann M et al. Disruption of the murine IL-4 gene blocks Th₂ cytokine responses. *Nature*, 1993, 362: 245~ 248.
 - 29 Pene J, Rousset F, Briner F et al. Interleukin-5 enhances interleukin 4-dependent IgE production by normal human B cells: the role of soluble CD 23 antigen. *Eur J Immunol*, 1988, 18: 929~ 935.
 - 30 Chu H W, Wang J M, Boutet M et al. Immunohistochemical detection of GM-CSF, IL-4 and IL-5 in a murine model of allergic bronchopulmonary aspergillosis. *Clin Exp Allergy*, 1996, 26: 461~ 468.
 - 31 Coffman R L, Seymour B W P, Hudak S et al. Antibody to interleukin-5 inhibits helminth-induced eosinophilia in mice. *Science*, 1989, 246: 308~ 310.
 - 32 Wardlaw A J, Dunnette S, Gleich G J et al. Eosinophils and mast cells in bronchoalveolar lavage in subjects with asthma: relationship to bronchial hyperreactivity. *Am Rev Respir Dis*, 1988, 137: 62~ 69.
 - 33 Gundel R H, Letts L G, Gleich G J. Human eosinophil major basic protein induces airway constriction and airway hyperresponsiveness in primates. *J Clin Invest*, 1991, 87: 1470~ 1473.
 - 34 Lefort J, Nahori M A, Ruffie C et al. In vivo neutralization of eosinophil-derived major basic protein inhibits antigen-induced bronchial hyperreactivity in sensitized guinea pigs. *J Clin Invest*, 1996, 97: 1117~ 1121.
 - 35 Busse W W, Calhoun W F, Sedgwick J D. Mechanism of airway inflammation in asthma. *Am Rev Respir Dis*, 1993, 147: S20~ S4.

(责任编辑: 蒋汉明 邓大玉)

1998年度国家自然科学基金概况

国家自然科学基金委员会 1998年度批准面上项目 3 552项,资助总经费 43 530万元,重点项目 175项,资助总经费 15 733万元;高技术新概念新构思探索项目 140项,资助总经费 1 763万元;高技术新概念新构思探索重点项目 17项,资助总经费 1 430万元。1998年度批准的面上项目比 1997年增加 98项,资助经费增加 4 028万元,平均资助强度从去年的 11.47元/项增加至 12.37元/项;项目数的批准率为 19.0%,比 1997年的 17.7%略高。获得批准的面上项目按类型划分为:自由申请为 2 775项,青年基金 631项,地区基金 146项。按学部划分为:数理科学部 475项,化学科学部 390项,生命科学部 1 240项,地球科学部 403项,工程与材料科学部 551项,信息科学部 359项,管理科学部 143项。

(摘自中国科学院 1999年《科学发展报告》P224)

Guangxi Sciences, Vol. 6 No. 1, February 1999